Chart I

Stereochemistry of Palytoxin. 1. C85-C115 Segment[†]

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Palytoxin, the toxic principle isolated from marine soft corals of the genus Palythoa, is the most poisonous substance known to date except for a few polypeptides and proteins found in bacteria and plants. Pioneering investigations by the Hawaii group¹ and by the Nagoya group² have recently led them independently to suggest the gross structure of palytoxin.³ In this series of papers, we will describe the complete assignment of the stereochemistry of palytoxin⁴ primarily on the basis of organic synthesis.

We chose to study degradation product $1a^{2d,1a}$ as our first target.



The structure, including the absolute configuration, of acetal 2b, which is a more advanced degradation product of 1b, was determined by Hirata, Uemura, and their co-workers using X-ray analysis.^{2a,c} A stereospecific, practical synthesis of 2a via 3 and 4 was recently achieved in our laboratory.⁵ We next turned our attention to the degradation product 1a, which had two unassigned

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(1) (a) Moore, R. E.; Bartolini, G. J. Am. Chem. Soc. 1981, 103, 2491.
(b) Moore, R. E.; Woolard, F. X.; Bartolini, G. Ibid. 1980, 102, 7370.
(c) Moore, R. E.; Woolard, F. X.; Sheikh, M. Y.; Scheuer, P. J. Ibid. 1978, 100, 7758.
(d) Moore, R. E.; Dietrich, R. E.; Hatton, B.; Higa, T.; Scheuer, P. J. J. Org. Chem. 1975, 40, 540.
(e) Moore, R. E.; Scheuer, P. J. Science (Washington, D.C.) 1971, 172, 495.
(2) (a) Uemura. D.; Ueda. K.: Hirata V: Naoki, H.; Washita, T. Tat-

(Washington, D.C.) 1971, 172, 495.
(2) (a) Uemura, D.; Ueda, K.; Hirata, Y.; Naoki, H.; Iwashita, T. Tetrahedron Lett. 1981, 22, 2781. (b) Uemura, D.; Ueda, K.; Hirata, Y.; Naoki, H.; Iwashita, T. Ibid. 1981, 22, 1909. (c) Uemura, D.; Ueda, K.; Hirata, Y.; Katayama, C.; Tanaka, J. Ibid. 1980, 21, 4861. (d) Uemura, D.; Ueda, K.; Hirata, Y.; Katayama, C.; Ueda, K.; Hirata, Y. John, S. J. Ueda, K.; Hirata, Y. J. John, S. J. Uemura, D.; Ueda, K.; Hirata, Y.; Katayama, C.; Tanaka, J. Ibid. 1980, 21, 4861. (d) Uemura, D.; Ueda, K.; Hirata, Y. J. Am. Chem. Soc. 1980, 102, 875.
(f) Hirata, Y.; Uemura, D.; Ueda, K.; Takano, S. Pure Appl. Chem. 1979, 51, 1975. *51*, 1875.

(3) For the structure and numbering of palytoxin, see part 4 of this series. (4) Palytoxin used in this study was extracted from Okinawan Palythoa tuberculosa. See ref 2 and also part 4 of this series. (5) Ko, S. S.; Klein, L. L.; Pfaff, K.-P.; Kishi, Y. Tetrahedron Lett. 1982,

23, 4415.



asymmetric centers. In order to determine the configuration of these asymmetric centers unambiguously, we decided to synthesize all the possible stereoisomers of 1a or its equivalent. Using the carbohydrate chain-extension method developed in our laboratory,6 we synthesized the two erythro isomers 6^7 and 7 (Chart I) from trans-allylic alcohol 5, which was obtained from 4. The assignment of the absolute configuration of these compounds depended upon Sharpless' asymmetric epoxidation,8 which is reliable for transallylic alcohols.9 Since the stereochemical outcome of Sharpless' asymmetric epoxidation of cis-allylic alcohols is not always predictable,⁹ the two three alcohols 8 and 9 were prepared from the intermediates used in the transformation of 5 into 6 and $7.^{10}$ The chemical shifts and spin-spin coupling constants for the C99, C100, and C101 protons of only isomer 9 were found to closely resemble those of 1a, suggesting that 9 possesses the natural

⁽⁶⁾ Minami, N.; Ko, S. S.; Kishi, Y. J. Am. Chem. Soc. 1982, 104, 1109. Finan, J. M.; Kishi, Y. Tetrahedron Lett. 1982, 23, 2719. Similar methods Finan, J. M.; Kishi, Y. Tetrahedron Lett. 1982, 23, 2719. Similar methods were also reported by Corey (Corey, E. J.; Hopkins, P. B.; Murnee, J. E.; Marfat, A.; Hashimoto, S. J. Am. Chem. Soc. 1980, 102, 7986), Roush (Roush, W. R.; Brown, R. J. J. Org. Chem. 1982, 47, 1371), and Masamune and Sharpless (Katsuki, T.; Lee, A. W. M.; Ma, P.; Martin, V. S.; Masamune, S.; Sharpless, K. B.; Tuddenham, D.; Walker, F. J. Ibid. 1982, 47, 1373. Ma, P.; Martin, V. S.; Masamune, S.; Sharpless, K. B.; Viti, S. M. Ibid. 1982, 47, 1378. Lee, A. W. M.; Martin, V. S.; Masamune, S.; Sharpless, K. B.; Walker, F. L. J. Am. Chem. Soc. 1982, 104, 3515) F. J. J. Am. Chem. Soc. 1982, 104, 3515).

⁽⁷⁾ Satisfactory spectroscopic data were obtained for all new compounds

⁽⁸⁾ Katsuki, T.; Sharpless, K. B. J. Am. Chem. Soc. 1980, 102, 5974. The latest publication on this subject from the Sharpless group is given in the following: Martin, V. S.; Woodard, S. S.; Katsuki, T.; Yamada, Y.; Ikeda, M.; Sharpless, K. B. Ibid. 1981, 103, 6237.

⁽⁹⁾ For examples, see: Nagaoka, H.; Kishi, Y. *Tetrahedron* 1981, 37, 3873. See also the references cited in footnote 6.

⁽¹⁰⁾ This was performed by the following steps: (1) Swern oxidation of five-membered carbonate alcohols (cf. compound 18 in our J. Am. Chem. Soc. publication cited under footnote 6); (2) $NaBH_4$ or $Zn(BH_4)_2$ reduction; (3) TLC separation; (4) aqueous base hydrolysis. For an alternative solution, see the last Masumune-Sharpless publication cited under ref 6.

configuration at these asymmetric centers. Indeed, the tetraacetate 1a [¹H NMR (CDCl₃) δ 2.03 (3 H, s), 2.07 (3 H, s), 2.09 (3 H, s), 2.10 (3 H, s); $\alpha_{\rm D}$ + 68.8° (c 0.52, CHCl₃)], synthesized from 3 and a derivative of 9,¹¹ was found to be identical with degradation product 1a¹² on comparison of spectroscopic data, establishing the stereochemistry of palytoxin at C100 and C101 as shown in 1a

We next turned our attention to nonaacetate 10,13 a degradation



product known to contain the C84-C98 carbon backbone.^{2d,1a} The ¹H NMR spectrum of 10 suggested that the relative stereochemistry of the tetrahydropyran portion was as indicated in the structure.^{1a,2d} However, the relative stereochemistry of the acyclic portion remained unknown. To determine the relative and absolute sterochemistry of 10 efficiently, we initially studied the more advanced degradation product 11.14

3,4,6-Tribenzyl-D-mannose 1,2-epoxide (12)15 was reacted with



the Grignard reagent prepared from (S)-(+)-3-tert-butoxy-2methyl-1-bromopropane¹⁶ in the presence of Li₂CuCl₄¹⁷ to yield stereoselectively alcohol 13. Swern oxidation¹⁸ of 13 followed by diborane reduction¹⁹ furnished an 8:1 mixture of alcohols 14 and 13. Debutylation of 14 followed by debenzylation and acetylation gave pentaacetate 11 [¹H NMR (CDCl₃) δ 0.96 (3 H, d, J = 7 Hz), 2.03 (3 H, s), 2.04 (3 H, s), 2.06 (3 H, s), 2.08 (6 H, s); $\alpha_{\rm D}$ +64.7° (c 0.65, CHCl₃)]. Starting with 12 and (R)-(-)-3tert-butoxy-2-methyl-1-bromopropane,16 the same sequence of reactions produced the C91 diastereomer of 11. On comparison of the spectroscopic data and optical rotations, synthetic pentaChart II



acetate 11 was found to be identical with degradation product 11, establishing the stereochemistry at C91, C93, C94, C95, C96, and C97.

In order to study the stereochemistry at C88, C89, and C90, we converted 14 to trans-allylic alcohol 15 (Chart II) and then subjected it to the carbohydrate chain-extension method⁶ in a manner identical with that used for allylic alcohol 5. The triols 16-19 resulted. The three relationship of triols 18 and 19, and consequently the erythro relationship of triols 16 and 17, was further confirmed by the fact that OsO4 oxidation of the tertbutyldiphenylsilyl ether of trans-allylic alcohol 15 followed by $(n-Bu)_4$ NF treatment furnished a 1:1 mixture of triols 18 and 19.

The acetonide alcohol 20, prepared from a derivative of threo-triol 19,²⁰ was subjected to Swern oxidation¹⁸ followed by addition of 3-butenylmagnesium bromide to the resulting aldehyde to yield approximately a 1:2 mixture of alcohols 21 and 22.²¹ Osmium tetroxide oxidation of the minor alcohol 21 followed by aqueous acid hydrolysis, debenzylation, and acetylation furnished nonaacetate 10. The same sequence of reactions on 22 gave the corresponding nonaacetate. On comparison of the spectroscopic data, synthetic nonaacetate 10²² was found to be identical with degradation product 10,²² while the nonaacetate derived from 22 was different. The corresponding nonaacetates were likewise synthesized from derivatives of the remaining triols 16-18, and none was found to be identical with degradation product 10.

H)CH₂OH in structure 9, was first converted to the pivaloyl acetonide [(1) (Me)₃CCOCl/py and (2) MeC(OMe)₂Me/acetone/p-TSA] and then subjected to the following sequence of reactions: (1) (n-Bu)₄NF/THF; (2) Swern oxidation; (3) Wittig reaction with 3 (LDA/DMF-THF); (4) HN=NH (5) $H_2/Pd-C/MeOH$; (6) aqueous NaOH; (7) aqueous AcOH; (8) Ac_2O/py .

⁽¹²⁾ Degradation product 1a was prepared by hydrogenolysis (Pd-C/AcOH-MeOH) of 1b.^{2d,la}

⁽¹³⁾ This substance was a diastereomeric mixture at C85. In this series of papers a small circle such as the one in structure 10 indicates that a degradation product (or its related compound) is a diastereomeric mixture due to the asymmetric center introduced during degradation reactions.

⁽¹⁴⁾ This substances was prepared from 10 in three steps: (1) aqueous NaOH; (2) NaIO₄/MeOH, followed by NaBH₄ workup; (3) Ac₂O/py,

followed by silica gel TLC separation. (15) Sondheimer, S. J.; Yamaguchi, H.; Schuerch, C. Carbohydr. Res. 1979, 74, 327.

⁽¹⁶⁾ This substance was prepared from (S)-(+)-3-hydroxy-2-methylpropionic acid according to the Cohen-Saucy procedure (Cohen, N.; Eichel, W. F.; Lopresti, R. J.; Neukom, C.; Saucy, G. J. Org. Chem. 1976, 41, 3505) We are indebted to Dr. Cohen for a generous gift of the acid.

 ⁽¹⁷⁾ Tamura, M.; Kochi, J. Synthesis 1971, 303.
 (18) Mancuso, A. J.; Huang, S.-L.; Swern, D. J. Org. Chem. 1978, 43. 2480

⁽¹⁹⁾ Garegg, P. J.; Maron, L. Acta Chem. Scand., Ser. B 1979, 33B, 453.

⁽²⁰⁾ The acetonide alcohol 20 was synthesized from 19 in three steps: (1) (Me)₃CCOCl/py; (2) MeC(OMe)₂Me/acetone/p-TSA; (3) LiAlH₄/THF.

⁽²¹⁾ The stereochemistry outcome of this reaction is not necessarily surprising; for examples see: Suzuki, K.; Yuki, Y.; Mukaiyama, T. Chem. Lett. 1981, 1529 and references cited therein. Bernardi, R.; Fuganti, C.; Grasselli, P. Tetrahedron Lett. 1981, 22, 4021 and references cited therein. Danishefsky, S.; Kobayashi, S.; Kerwin, J. F., Jr. J. Org. Chem. 1982, 47, 1981 and references cited therein.

⁽²²⁾ This substance was a diastereomeric mixture due to the C85 position. Separation of the diastereomers was possible by preparative silica gel TLC Separation of the diastereomers was possible by preparative since get 1LC (Merck silica get 5769; solvent system 5:1 ether-hexane). The less polar nonaacetate **10** had the following spectroscopic data: ¹H NMR (CDCl₃) δ 0.93 (3 H, d, J = 7 Hz), 2.02 (6 H, s), 2.07 (3 H, s), 2.08 (3 H, s), 2.09 (3 H, s), 2.09 (6 H, s), 2.10 (3 H, s), 2.11 (3 H, s); α_D +36.2° (*c* 0.41, CHCl₃). The more polar nonaacetate **10** had the following: ¹H NMR (CDCl₃) δ 0.93 (3 H, d, J = 7 Hz), 2.03 (3 H, s), 2.04 (3 H, s), 2.06 (3 H, s), 2.07 (6 H, s), 2.09 (6 H, s), 2.10 (3 H, s), 2.14 (3 H, s); α_D +31.1° (*c* 0.13, CHCl₃).

The stereochemistry of alcohols 21 and 22 was established by the following experiments. Benzylation of the major alcohol 22 followed by aqueous acid hydrolysis, periodate oxidation, and sodium borohydride reduction afforded 2-(benzyloxy)hex-5-en-1-ol (23). The α_D value of 23 was found to be -9.4°. The specific



rotations of (R)- and (S)-2-(benzyloxy)hex-5-en-1-ols, prepared from D- and L-glyceraldehyde ketals,²³ were found to be -11.7and $+10.4^{\circ}$, respectively. Thus, C88 has the S configuration in the minor alcohol 21. This conclusion was further confirmed by comparison of the ¹H NMR spectra of MTPA²⁵ esters of 23 obtained from the above-mentioned sources.

The experiments summarized above allowed us to assign the stereochemistry of degradation product 10 as indicated. Since this assignment of the relative stereochemistry between C90 and C91 was based solely on the results of Sharpless' asymmetric epoxidation⁸ of 15, we felt it was desirable to have additional evidence.²⁶ For this reason, the stereochemistry assignment at C88, C89, and C90 was performed by an alternative method. Thus, cis-allylic alcohols 24 were prepared from 14²⁴ and subjected to OsO₄ oxidation, aqueous acid hydrolysis, debenzylation, and acetylation to yield a mixture of nonaacetates corresponding to 10. However, none of these nonaacetates was identical with degradation product 10, establishing the relative stereochemistry at C88 and C89 as threo. This information, along with knowledge of the relative stereochemistry between C89 and C90 and of the absolute stereochemistry at C88 and C91 (vide supra), excluded all possible structures for degradation product 10 except the one shown.

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Supplementary Material Available: Spectroscopic data for compounds 1a, 10 (two diastereomers), and 11 and details of some synthetic sequences (2 pages). Ordering information is given on any current masthead page.

Stereochemistry of Palytoxin. 2.1 C1-C6, C47-C74, and C77-C83 Segments[†]

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Continuing from the preceding communication, we will describe the stereochemistry assignment of the C1-C6, C47-C74, and C77-C83 portions of the marine natural product palytoxin.²

The lactone diacetate 1 (Chart I), containing C1-C6, is a known degradation product of palytoxin. The ¹H NMR spectrum of 1 suggested that the relative stereochemistry at C2, C3, and C5 was as indicated in 1, but the absolute stereochemistry was unknown.³ By use of the carbohydrate chain-extension method,⁴ tetraacetate 2^{5} [¹H NMR (CDCl₃) δ 0.97 (3 H, d, J = 6.9 Hz), 2.05 (3 H, s), 2.06 (3 H, s), 2.07 (3 H, s), 2.09 (3 H, s); $\alpha_{\rm D}$ +17.0° (c 0.17, $CHCl_3$] was synthesized from (S)-(+)-3-hydroxy-2-methylpropionic acid.^{6,7} Upon comparison of spectroscopic data and optical rotations, tetraacetate 2 was found to be identical with the tetraacetate prepared from the degradation product,⁸ establishing the absolute stereochemistry of C2, C3, and C5.

The tetraacetate 3, containing C47-C56, was isolated as a degradation product of palytoxin.⁹ By use of the carbohydrate chain-extension method,⁴ triacetate 4 [¹H NMR (CDCl₃) δ 0.95 (3 H, d, J = 7.4 Hz), 2.02 (3 H, s), 2.03 (3 H, s), 2.04 (3 H, s); $\alpha_{\rm D}$ -14.2° (c 0.73, CH₂Cl₂)] and its C49 diastereomer were synthesized from (S)-(+)-3-hydroxy-2-methylpropionic acid.^{6,7} On comparison of spectroscopic data and optical rotations, triacetate 4 was found to be identical with advanced degradation product 4^{10} Wittig reaction of aldehyde 6^7 with phosphonium salt 5⁷ followed by debenzylation and acetylation gave trans-olefin **3** [¹H NMR (CDCl₃) δ 1.01 (3 H, d, J = 7.0 Hz), 2.03 (3 H, s), 2.04 (3 H, s), 2.05 (6 H, s)]. Starting with 5 and the antipode⁷ of 6, the C53 diastereomer of 3 was also prepared. Upon comparison of ¹H NMR data, synthetic tetraacetate 3 was found to be identical with degradation product 3, establishing the stereochemistry at C49, C50, and C53.

The pentaacetate 7, which contains C77-C83, is a degradation product of palytoxin. The relative stereochemistry of 7 was found

- (5) Satisfactory spectroscopic data were obtained for all new compounds in this paper. (6) We are indebted to Dr. Cohen, Hoffmann-La Roche Inc., for a gen-
- erous gift of this acid.
 - (7) Details of this synthesis are given in the supplementary material.
- (8) This substance was prepared from the degradation product reported as compound 1 in ref 2f of the preceding paper in four steps: (1) $O_3/$ MeOH/-78 °C, followed by NaBH₄ workup; (2) Ac₂O/py; (3) LiAlH₄/ THF/0 °C; (4) Ac₂O/py
 - (9) See ref 2f and 2d in the preceding paper.

⁽²³⁾ D-Glyceraldehyde acetonide was prepared according to the method (25) D-Orden deriver action was prepared actioning to the intention reported in the following: Fischer, H. O. L.; Baer, E. *Helv. Chim. Acta* 1934, 17, 622). With the procedure reported for the D-series (Zinner, H.; Milbradt, J. *Carbohydr. Res.* 1966, 2, 470), L-glyceraldehyde cyclohexanone ketal was prepared from L-arabinose. Transformation of glyceraldehyde ketals into 23 was achieved in cited tester 24 was achieved in eight steps.24

⁽²⁴⁾ Details of this synthesis are given in the supplementary material. (25) Dale, J. A.; Dull, D. L.; Mosher, H. S. J. Org. Chem. 1969, 34, 2543. Sullivan, G. R.; Dale, J. A.; Mosher, H. S. Ibid. 1973, 38, 2143

⁽²⁶⁾ Chemical correlation of optically active epoxides prepared by asymmetric epoxidation has been performed in many cases; for example, see footnotes 6, 8, and 9.

⁺This work was presented by Y. Kishi as part of a lecture at the symposium honoring the memory of Dr. Willy Leimgruber on March 26, 1982, at Rutgers University, Newark, NJ.

⁽¹⁾ Part 1 of this series: J. Am. Chem. Soc. preceding paper in this issue. (2) For the structure and numbering of palytoxin, see part 4 of this series.

⁽³⁾ This assignment was made based on the spin-spin coupling constants $J_{2,3} = 11.6$ Hz and $J_{4,5} = 12.0$ and 3.5 Hz, given in the supplementary material

for ref 1a of the preceding paper. (4) See ref 6 in the preceding paper.

⁽¹⁰⁾ This substance was prepared from the degradation product reported as compound **21** in ref 1a of the preceding paper in three steps: (1) NaOH/MeOH/room temperature; (2) $NaIO_4/H_2O/0$ °C, followed by NaBH₄ workup: (3) Ac_2O/py , followed by TLC separation.